### **Chapter 2: Buffers and Titrations**

#### **Purpose:**

1) Get to know your pH meter

2) Make a common buffer used in biochemistry and perform titrations of that buffer with acid or base to find the pK<sub>a</sub> values for the buffer

3) Hydrolyze BSA with trypsin and calculate the number of Lys and Arg residues that BSA contains



# pH Meter

- Glass-electrode sensitive to hydrogen ions
- Electrode somewhat sensitive to other alkali metals
- Complete system contains:
  - Electrometer 5
  - Reference Electrode 6
  - Solution to be measured 1,4
  - Glass Electrode 2,3

# Titration Curves in Non-buffered Solutions

Weak Acid = 0.1 M Acetic Acid Strong Acid = 0.1 M Hydrochloric Acid



- Equivalence Point
  - Point at which reaction is neutralized
  - Inflection point in titration curve
- Strong Acid pH 7.0
- Weak Acid pH 8.8
- Buffered solutions behave as weak acids
- Table of pK<sub>a</sub> values Lab Manual p. 36

#### pH Changes in Buffered Solutions



#### **Buffered Titration Curve**



#### **Buffered Titration Curve**



- Empirically, H-H equation useful for buffering range
- Buffers most effective near pK<sub>a</sub>

# $pH = pK_a$ when $[A^-] = [HA]$

## **Buffering Capacity**

 Ability of buffer to resist changes in pH with addition of acid or base

Buffer Capacity = 
$$\frac{-dH^+}{dpH}$$
 = 2.303  $\left[\frac{[A^-][HA]}{[A^-]+[HA]}\right]$ 

 Highest buffering capacity obtained when [A<sup>-</sup>] = [HA]

#### **Procedure:** Titration

- Make His Buffer
  - Starting pH?
- Four Titrations
  - Titrate Acid Group of His
  - Titrate the Two Basic Groups of His
  - Titrate Water with Acid
  - Titrate Water with Base
- Subtract Water Values from His to Get Pure His Curve



#### **Procedure:** Titration

- Make His Buffer 0.4 M His-HCl = 0.4 M HA
- Deprotonated His (His<sup>0</sup>) =  $[A^-] = [H^+]$



#### **Procedure:** Titration

- Make His Buffer
  - Starting pH = 3.22
- Four Titrations
  - Titrate Acid Group of His
  - Titrate the Two Basic Groups of His
  - Titrate Water with Acid
  - Titrate Water with Base
- Subtract Water Values from His to Get Pure His Curve



#### Digestion of BSA with Trypsin

#### **Proteolytic Cleavage of Proteins**

Trypsin

Cleaves C-terminal of (+) charged side chains

### Trypsin



Cengage Learning

# Procedure: Determining the Number of Lys and Arg (combined) in BSA

- Denature BSA at 80-90 °C until cloudy
- Digest BSA with Trypsin
  - Titrate during reaction to maintain pH value 8.5
  - Indicate volume KOH added and the time elapsed
- Calculate the Number of Peptide Bonds Cleaved
  When Reaction is Complete
  - Calculate mmols KOH added at endpoint
  - Calculate number of Arg + Lys per molecule BSA



**New N-Termini Add to Buffer Capacity** 

- Since pH is only slightly greater than the pK<sub>a</sub> of Nterminus
  - Each new N-terminus will buffer the new H<sup>+</sup> released from the reaction
    - Not every amino group will gain a proton
- How much H<sup>+</sup> is actually produced?
  - Depends on ratio of [A<sup>-</sup>]/[HA]
  - If pH is constant, [A<sup>-</sup>]/[HA] must remain constant

Problem 10, p. 43: What is ratio of [A<sup>-</sup>]/[HA] for the protonation of an amine with a pK<sub>a</sub> = 8.2, at pH 8.5?

$$R-NH_{2} + H^{+} \iff R-NH_{3}$$
$$pH = pK_{a} + \log \frac{[A^{-}]}{[HA]}$$

 $8.5 = 8.2 + \log [R-NH_2]/[R-NH_3] = 8.2 + \log [A^-]/[HA]$ 

pH of Amino reaction group  $pK_a$  0.3 = log [A<sup>-</sup>]/[HA] [A<sup>-</sup>]/[HA] = 10<sup>0.3</sup> = 2/1

2/3 depronated [A<sup>-</sup>], 1/3 protonated [HA]

- The trypsin digestion alters the buffer capacity of the solution
  - As more amino groups are formed, some accept a proton
  - Other protons are neutralized by KOH titration
- Total # of peptide bonds cleaved = (mmol of KOH added)(3 peptide bonds cleaved/2 mmol KOH added)
- Total # of Lys + Arg per molecule of BSA = (# of peptide bonds cleaved)/(mmol of BSA used)
  - Calculate mmol of BSA using MW (66,000 g/mol)